

Single and Joint Toxicity of Sulfamonomethoxine and Cadmium on Three Agricultural Crops

CAIXIA JIN,¹ JING FAN,¹ RUI LIU,² AND RUILIAN SUN³

¹Key Laboratory for Yellow River and Huai River Water Environment and Pollution Control, Ministry of Education, Henan Key Laboratory of Environmental Pollution Control, Henan Normal University, Xinxiang, China ²Key Laboratory of Pollution Ecology and Environmental Engineering, Institute of Applied Ecology, Chinese Academy of Sciences, Shenyang, China ³Environment Research Institute, Shandong University, Jinan, Shandong, China

In order to evaluate ecological risk of agrochemicals in an agricultural environment, single and joint toxic effects of sulfamonomethoxine (SMM) and cadmium (Cd) on seed germination of wheat (Triticum aestivum L.), Chinese cabbage (Solanum lycopersicum), and radish (Raphanus sativus L.) were investigated. The results showed that the root and shoot elongation were significantly affected under the contamination condition (P < 0.05). In the single-factor experiments, the toxic effect of SMM on crops was much stronger than that of Cd. In the joint effects of SMM and Cd, the interactive effects of the two pollutants on root/shoot elongation of the three crops were complicated, while SMM and Cd had significantly (P < 0.05) synergic effects on root/shoot elongation of SMM, SMM and Cd had significantly (P < 0.05) synergic effects on root/shoot elongation of SMM, some and Cd had significantly (P < 0.05) synergic effects on root/shoot elongation of SMM, some and Cd had significantly (P < 0.05) synergic effects on root/shoot elongation of SMM, some and Cd had significantly (P < 0.05) synergic effects on root/shoot elongation of synergic effects on root/shoot elongation of the three crops, but antagonistic effects were found when the concentration of SMM was high.

Keywords Sulfamonomethoxine, cadmium, joint toxicity, inhibitory rate, crops

Introduction

Veterinary antibiotics are physiologically highly active substances and are used on a large scale as therapeutic drugs and feed additives in modern agricultural practice for the prevention and treatment of bacteria-borne diseases and improved growth rates (Boxall *et al.*, 2002; Jørgensen and Halling-Sørensen, 2000; Klaus, 2009a; 2009b). But it has been reported that only a fraction of the drugs are absorbed by animals after oral administration; a large percentage are excreted and released into the environment; for example, via manure, sludge, and waste water used as fertilizer or irrigation water in agricultural lands (Jørgensen and Halling-Sørensen, 2000; Kong and Zhu, 2007; Nicole, 2008; Wei *et al.*, 2011). The residual concentration of veterinary drugs in soil has ranged from $\mu g/$ kg to g/kg (Kong and Zhu, 2007); this poses an important potential threat to the ecosystem and to human health. Therefore, the effect of residual veterinary drugs on the ecosystem is a growing environmental concern (Boxall *et al.*, 2002; Thiele-Bruhn, 2003).

Sulfonamides, as one of the selective veterinary antibiotics for human and animal

Address correspondence to Rui Liu, Institute of Applied Ecology, Chinese Academy of Sciences, No. 72, Wenhua Road, Shenyang 110016, Liaoning, China. E-mail: liurui@iae.ac.cn



treatment, are widely used for the control of most gram-positive and gram-negative microorganisms by inhibiting the multiplication of bacteria as competitive inhibitors of p-aminobenzoic acid in the folic acid metabolism cycle (Accinelli et al., 2007; Baran et al., 2006). In the 1990s, a survey in several countries revealed that sulfonamides account for 11–23% of the applied veterinary antibiotics in the European Union (Thiele-Bruhn, 2003). According to Thiele-Bruhn et al. (2004), as much as 90% of sulfonamides are excreted after consumption. Some research has indicated that sulfonamides would not be easily degradable in a soil environment (Ingerslev and Halling-Srensen, 2000; Holger et al., 2008). In the past several years, sulfonamides have been widely detected in manure, soils, and surface waters (Boxall et al., 2003; Christian et al., 2003; Nicole, 2008). Other investigations have also demonstrated the potential for sulfonamides to contaminate subsoil (Boxall et al., 2002). Like other veterinary antibiotics, sulfamonomethoxine (SMM), a very common sulfonamide drug, entered agricultural soils mainly by fertilizing, grazing (Dagnac et al., 2002; Haller et al., 2002; Jørgensen and Halling-Sørensen, 2000), transportation through soil (Christian et al., 2003; Hamscher et al., 2005; Wojciech et al., 2006), surface runoff (Davis et al., 2006), and uptake by plants (Dolliver et al., 2007).

On the other hand, cadmium (Cd) is widely used in modern industry. A total of 2.2×10^4 tons of Cd has been discharged into the environment during the past half-century (Singh *et al.*, 2003). According to Zhou *et al.* (2004a), the concentration of Cd in some severely contaminated soils (a result of mining, smelting, and wastewater irrigation) was higher than 100 mg·kg⁻¹. Since Cd is not degradable, the accumulation of Cd in the soil may have durable effects on the plants growing on the soil. As a result, the pollution of water-soil-plant systems is becoming the focus of ecological studies (Zhou, 2003; Wang and Zhou, 2006). It is not difficult to understand that there is more frequent coexistence of SMM and Cd in agricultural lands. Increasing attention has been paid to the pollution problems associated with sulfonamides and Cd, respectively; however, little information is available about the joint toxic effects of SMM and Cd.

Seed germination and root elongation tests have been used as acute phytotoxicity tests to provide valuable information about the toxicity of contamination in the environment and other parameters (Liu *et al.*, 2009). This test has several advantages (sensitivity, simplicity, low cost, and suitability for unstable chemicals or samples) over other toxicity tests and is suitable as a standby test method as well as a rapid tool to evaluate the ecological risk of hazardous chemicals (Wang *et al.*, 2002; Lin and Xing, 2007). Thus, in this study, wheat, radish, and Chinese cabbage, the main crops in north China, were chosen as target testing plants to examine the single and joint effects of SMM and Cd on seed germination and shoot and root elongation.

Materials and Methods

Materials

All reagents used in this study were of analytic grade. The tested form of cadmium was $CdCl_2 \cdot 2.5H_2O$. Sulfamonomethoxine (SMM) was obtained from Boyahua Science and Technology Company in Beijing, China, and its type is 92.9% of dispersible granule. The molecular formula of SMM is $C_{11}H_{12}N_4O_3S$ and its structure is shown in Figure 1.

The seeds of wheat (*Triticum aestivum L.*), Chinese cabbage (*Solanum lycopersicum*), and radish (*Raphanus sativus L.*) were purchased from Seed Corporation of Xinxiang City. The crop seeds were immersed in 10% sodium hypochlorite solution for





Figure 1. Molecular structure of SMM.

10 min to ensure sterility, and then they were washed several times with deionized water. The surface soil samples (0–20 cm) were collected from an experimental plot at the College of Life Sciences, Henan Normal University ($35^{\circ}31'N$ and $113^{\circ}85'E$), Xinxiang, Henan province, China, which had not been fertilized for almost 30 years. The fresh soil samples were air-dried and ground to pass a sieve of 1.0 mm before use. The physicochemical prosperities of the soil were: pH, 7.83; OM, 1.65%; CEC, 12.26 cmol/kg; total N, 0.09 mg·kg⁻¹; total P, 0.04 mg·kg⁻¹; total K, 0.18 mg·kg⁻¹; total Cd, 0.066 mg·kg⁻¹.

Toxicity Experiments

456

A 50 g sample amended with different concentrations of tested pollutions (SMM and Cd) was put into a culture dish. The soil moisture was adjusted with deionized water until it reached 60% of maximal holding capacity, and then mixed well. Fifteen sterilized seeds of three crops were scattered with a tweezer in soil in a culture dish and covered, then the dishes were put into the culturing box (HG-303-3, made in Nanjing, China) under dark conditions with temperature being controlled at $25 \pm 1^{\circ}$ C. All treatments were replicated three times to minimize experimental errors. When the length of the growing root cultured in the control soil without SMM and Cd reached 20 mm (December 13, 2012), the exposed experiment was finished, and the seed germination and the shoot and root elongation of all of the treatments were measured and calculated.

Single-Factor Experiments

According to 10–50% of the root elongation inhibitory rate of three crops by Cd and SMM, the tested concentrations of Cd were 0, 100, 200, 300, 400, 500, 600 mg·kg⁻¹, and the tested concentrations of SMM were equal to 0, 2, 5, 10, 20, 40, and 60 mg·kg⁻¹ for wheat; 0, 0.5, 1, 2, 4, 6, and 8 mg·kg⁻¹ for Chinese cabbage; and 0, 0.5, 1, 2, 4, 8 and 16 mg·kg⁻¹ for radish.

Combined-Pollution Experiments

According to the results from the single-factor experiments, the tested concentration of SMM was 0, 2, 5, 10, 20, 40, and 60 mg·kg⁻¹ for wheat; 0, 0.5, 1, 2, 4, 6, and 8 mg·kg⁻¹ for Chinese cabbage; and 0, 0.5, 1, 2, 4, 8, and 16 mg·kg⁻¹ for radish. The tested



concentration of Cd was set to be 0, 200, and 500 $\text{mg}\cdot\text{kg}^{-1}$ in the joint effect experiment of SMM and Cd.

Data Reliability

The inhibition rate (IR,%) was calculated by Equation (1) as follows:

$$IR = \frac{A - B}{A} \times 100\%$$
 (1)

where A is the root or shoot length of contrast (mm) and B is root or shoot length of different concentration treatments (mm). Statistical analysis, including calculation of average values, standard deviation (S.D.), and regression, was performed on the data obtained in the tests with SPSS12.0. The multiple comparison procedure (LSR test) was used to compare shoot and root elongation under the combined pollution of SMM and Cd and statistical significance was set at P < 0.05.

Results

Toxic Effects of SMM on Three Crops

The statistical analysis showed that there were no significant differences (P > 0.05) in germination rate of the three crops (wheat, Chinese cabbage, and radish) exposed to the seven experimental concentrations of SMM, respectively. By contrast, SMM had a significantly (P < 0.05) toxic effect on root and shoot elongation of these crops under the experimental conditions (Figure 2). The inhibitory rate of root and shoot elongation of the three crops increased with the increasing concentration of SMM in soil. As shown in Figure 2, for each one of the three crops, there was a significant linear correlation between the inhibitory rate of root and shoot elongation and the tested concentration of SMM. The corresponding relationship can be expressed using the regression equations in Table 1. According to the regression equations based on the inhibition of root elongation and shoot elongation, IC_{50} (the concentration of SMM) corresponds to the 50% inhibitory rate of RI (the inhibitory rate (%) of root elongation) or SI (the inhibitory rate (%) of shoot elongation), mg kg^{-1}) of SMM was calculated. The calculation showed that IC₅₀ of SMM based on the inhibitory rates of root elongation of wheat, Chinese cabbage, and radish was 33.7, 4.41, and 8.82 mg kg⁻¹, respectively, which meant that the toxic effect of SMM on the three crops was in the following sequence: Chinese cabbage>radish>wheat. For IC_{50} of SMM based on the inhibitory rates of shoot elongation of the three crops, the order was the same. Thus, Chinese cabbage was the most sensitive to the toxicity of SMM.

As shown in Figure 2, under the same concentration of SMM, the inhibitory rate of root elongation of wheat was higher than that of shoot elongation; for example, when the concentration of SMM was 2 mg·kg⁻¹, the inhibitory rates of root elongation and shoot elongation were 9.6% and 3.4%, respectively; when the concentration of SMM was 20 mg·kg⁻¹, the inhibitory rates of root elongation and shoot elongation were 49.3% and 28.5%, respectively. The same results were found for Chinese cabbage and radish. The inhibitory effect on shoot elongation of SMM was weaker than that on root elongation, which meant that root was more sensitive than shoot under the pollution of SMM, while the statistical analysis showed that differences between root elongation and shoot





Figure 2. Toxic effects of SMM on the root and shoot elongation of the three crops (wheat, Chinese cabbage, and radish).

elongation were not significant (P > 0.05) at the high concentration of SMM. When the concentration of SMM in soil reached the highest for each crop in this experiment, the inhibitory rates of root elongation and shoot elongation of wheat, Chinese cabbage, and radish were 65.6% and 63.3%, 74.1% and 74.6%, 73.8%, and 73.1%, respectively.

Toxic Effects of Cd on Three Crops

Similar to the effects of SMM, root elongation and shoot elongation were correlated with the concentration of Cd (P < 0.05), and seed germination was not sensitive to Cd at the



458

Crop Species	Regression Equations ^a	\mathbb{R}^2	Р	IC ₅₀ ^b
Wheat	$RI = 0.8709C_{SMM} + 20.657$	0.8188	< 0.05	33.69
	$SI = 1.1012C_{SMM} + 5.9302$	0.9211	< 0.01	40.02
Chinese cabbage	$RI = 8.5443C_{SMM} + 12.331$	0.9263	< 0.01	4.41
-	$SI = 10.491C_{SMM} + 0.6209$	0.9461	< 0.01	4.71
Radish	$RI = 5.0153C_{SMM} + 5.7707$	0.8122	< 0.05	8.82
	$SI = 4.8835C_{SMM} + 2.9261$	0.8732	< 0.01	9.64

 Table 1

 Relationships between inhibitory rate of root elongation (RI) and shoot elongation (SI) of the three crops and the concentration of SMM in soil

 $^{a}C_{SMM}$ was the tested concentration of SMM, mg/kg.

^bIC₅₀ was the concentration of SMM correspond to 50% inhibitory rate of RI or SI, mg/kg.

tested concentration (P > 0.05). As shown in Figure 3, there were positive linear relationships between the inhibitory rate of root elongation, shoot elongation, and the concentration of Cd. The inhibitory rates of root and shoot elongation increased with increasing concentration of Cd in the tested soil. The corresponding regression equations are listed in Table 2. According to Table 2, Cd had obviously toxic effects on root and shoot elongation. Toxic effects on growth of the three crops were the strongest when the concentration of Cd was 600 mg·kg⁻¹; the inhibitory rates of root elongation of wheat, Chinese cabbage, and radish were 65.9%, 58.5% and 42.1%, respectively. According to Figure 3 and Table 2, it can be concluded that IC₅₀ of Cd based on the inhibition of root elongation of wheat, Chinese cabbage, and radish was equal to 507, 550, and 781 mg·kg⁻¹, respectively, which meant that the tolerance of radish was stronger than the other crops to the toxicity of Cd, and wheat was the most sensitive among the three crops.

As shown in Figure 3, when the concentration of Cd was 100 mg·kg⁻¹, the inhibitory rates of root elongation of wheat and radish were -4.2% and -6.1%, respectively. That means that Cd had a promoted effect on the growth of wheat and radish when the concentration of Cd was low in soil.

Toxic Effects of SMM and Cd on Three Crops

It was shown by the variance analysis that there were markedly positive correlations between the inhibitory rate of root and shoot elongation and the concentration of SMM when the concentration of Cd remained at 0, 200, and 500 mg·kg⁻¹, respectively (Figures 4 and 5). As shown in Table 3, SMM and Cd had synergic effects on the inhibition of root and shoot elongation of wheat. The root elongation of wheat was significantly reduced with increasing concentration of Cd in the tested soil at low concentrations of SMM (P < 0.05), which meant that the toxicity was mainly caused by Cd when the concentration of SMM was low. However, the synergic effects were not significant at high concentrations of SMM (P > 0.05). When SMM concentration was 40 mg·kg⁻¹ in soil and Cd was 0, 200, and 500 mg·kg⁻¹, respectively, the inhibitory rate of root elongation of wheat was 59.9%, 66.7%, and 69.2%, respectively.

The root and shoot elongation of Chinese cabbage and radish were also severely inhibited under the combined pollution of SMM and Cd (Tables 4 and 5). Joint effects of SMM and Cd on seeding growth of Chinese cabbage and radish showed a similar





Figure 3. Toxic effects of Cd on the root and shoot elongation of the three crops (wheat, Chinese cabbage, and radish).

tendency. As can be seen from Figures 4 and 5, about a 60% inhibitory rate was a dividing point. The synergic effect played a major role when the inhibitory rate was lower than 60%, but an antagonistic effect was observed when the inhibitory rate was higher than 60%. For example, when SMM concentration was 8 mg·kg⁻¹ in soil and Cd was 0, 200, and 500 mg·kg⁻¹, respectively, the inhibitory rate of root elongation of Chinese cabbage was 74.1%, 67.1%, and 66.1%, respectively. The inhibitory rate was gradually decreased with increasing concentration of Cd in the tested soil at the same concentration of SMM.

The corresponding regression equations can be expressed as those listed in Tables 6 and 7. Calculated from the regression equations, IC_{50} of SMM inhibiting root and shoot elongation of these crops was listed. It is revealed that the value of IC_{50} decreased



Table 2

Crop Species	Regression Equations ^a	R^2	Р	IC ₅₀ ^b
Wheat	$RI = 0.1284C_{Cd} - 15.134$	0.9319	< 0.01	507.27
	$SI = 0.1580C_{Cd} - 27.442$	0.9045	< 0.01	490.14
Chinese cabbage	$RI = 0.073C_{Cd} + 9.8104$	0.8438	< 0.01	550.54
C	$SI = 0.0894C_{Cd} + 9.1129$	0.9271	< 0.01	457.35
Radish	$RI = 0.0701C_{Cd} - 4.7733$	0.8122	< 0.05	781.36
	$SI = 0.0869C_{Cd} - 10.096$	0.9025	< 0.01	691.55

Relationships between inhibitory rate of root elongation (RI) and shoot elongation (SI) of the three crops and the concentration of Cd in soil

 ${}^{a}C_{Cd}$ was the tested concentration of SMM, mg/kg. ${}^{b}IC_{50}$ was the concentration of Cd correspond to 50% inhibitory rate of RI or SI, mg/kg.

obviously when the concentration of Cd remained at 0, 200, and 500 mg·kg⁻¹, as shown in Tables 6 and 7.

Discussion

In the single-factor experiment of SMM or Cd, seed germination of the three crops was not sensitive to the toxicity of the two pollutants. That may be explained by the protection of the seed coat. The seed coat plays a very important role in protecting the embryo from harmful external factors. Seed coats can have selective permeability (Malgorzata and Jolanta, 1998). Pollutants, though having an obviously inhibitory effect on root growth, may not affect germination if they cannot pass through seed coats. These results are consistent with those of Song et al. (2002) and Cheng and Zhou (2002). They identified, in their studies on the toxicity of heavy metals (Cd, Cu, Pb and Zn) and a chemical (X-3B red dye) reactive to wheat, that the inhibitory rate of root elongation was higher than that of germination rate at the same concentration of pollutants.

Cd inhibition on root elongation has been observed for some plants (Cheng and Zhou, 2002; Zhou, 2003). Cd could reduce ATPase activity of oat root, which hampered the absorption of potassium (K) by plant roots (He et al., 1998). The reason for this inhibition of root elongation may lie in the encumbrance of K absorption. However, many studies have confirmed that Cd could promote the growth of plants when the concentration of Cd is low (Lin et al., 2007; Aina et al., 2007; Robert and Joanna, 2003). In this study, Cd had a promoted effect on the root and shoot elongation of wheat and radish when Cd concentration was 100 mg kg^{-1} in soil. However, the mechanisms of stimulatory effects are not well understood (Lin et al., 2007). This phenomenon is normally related to a so-called "hormetic effect" that probably represents an "overcompensation" response to a disruption in the homeostasis of the organism (Aina et al., 2007). According to Robert and Joanna (2003), a low dose of Cd could stimulate cell proliferation, thus the growth of plants will be stimulated.

IC₅₀ was often used to evaluate the ecotoxic effects of the pollutants (Zhou et al., 2004a). Tables 1 and 2 illustrate that the effect of IC_{50} of SMM on root elongation of wheat, Chinese cabbage, and radish was 33.7, 4.41, and $8.82 \text{ mg} \cdot \text{kg}^{-1}$, respectively, whereas IC₅₀ of Cd was 507, 550, and 781 mg·kg⁻¹, respectively, which meant that the toxic effects of SMM on crops were much stronger than that of Cd. According to IC_{50} , different plants have





Figure 4. Joint toxic effects of SMM and Cd on root elongation of wheat, Chinese cabbage, and radish.

different sensitivities under the various pollutants. In this study, Chinese cabbage was the most sensitive to SMM and wheat was the most sensitive to the toxicity of Cd, which could be explained by the fact that different plants have their resistant mechanism to each kind of pollutant and the target molecule affected by each pollutant was different (Zhou *et al.*, 2004b). The mechanism of SMM on plants remains unknown; however, it would be closely related to the chemical composition and the nature of the molecules of SMM. Further research should focus on the specific effect mechanism.





Figure 5. Joint toxic effects of SMM and Cd on shoot elongation of wheat, Chinese cabbage, and radish.

The interactive effects of the SMM and Cd on root and shoot elongation of the three crops were very complicated. SMM and Cd had significantly (P < 0.05) synergic effects on root and shoot elongation of the three crops at the low concentration of SMM. However, the synergic effects were not significant (P > 0.05), and even antagonistic effects were found when the concentration of SMM was high, which meant the toxicity of SMM



لاستشارات کالا									
ił		Difference in	average root/shoo	t elongation (mear	Table 31, mm) of wheat u	nder joint pollutio	n of sulfamonomet	thoxine and cadmi	m
		Cd (mg/kg)			Sulfamonome	thoxine(mg/kg) (1	Mean \pm SD)		
			0	2	5	10	20	40	60
	root	0	26.32 ± 2.13	23.79 ± 3.16	20.64 ± 0.68	16.53 ± 2.05	13.34 ± 1.49	10.55 ± 0.92	9.05 ± 0.86
		200	20.83 ± 1.32	20.65 ± 2.81	16.47 ± 2.50	16.34 ± 1.76	11.53 ± 0.90	8.76 ± 0.63	6.73 ± 0.72
		500	13.24 ± 1.53	15.01 ± 1.42	14.23 ± 1.45	13.36 ± 1.83	10.34 ± 0.86	8.10 ± 0.74	5.92 ± 0.65
	shoot	0	8.13 ± 1.40	7.85 ± 1.24	7.36 ± 0.93	6.59 ± 0.89	5.81 ± 0.71	3.03 ± 0.63	2.99 ± 0.14
		200	7.40 ± 0.57	7.18 ± 1.45	5.92 ± 0.56	4.60 ± 0.84	3.02 ± 0.82	2.98 ± 0.58	2.87 ± 0.70
		500	3.17 ± 0.38	3.73 ± 0.96	3.65 ± 0.84	2.91 ± 0.79	2.84 ± 0.69	2.11 ± 0.42	2.31 ± 0.53

www.m

المستشارات		lifference in ave	rage Chinese cabb	age root/shoot elo	Table 4 ngation (mean, m	m) under joint pol	lution of sulfamor	nomethoxine and c	admium
		Cd (mg/kg)			Sulfamonome	thoxine(mg/kg) (]	Mean ± SD)		
			0	0.5	1	2	4	6	8
	root	0	31.33 ± 2.91	25.18 ± 0.76	26.72 ± 0.98	21.34 ± 1.15	18.18 ± 1.29	7.70 ± 1.52	8.13 ± 0.96
		200	21.77 ± 1.82	23.44 ± 1.45	20.33 ± 2.51	22.23 ± 1.26	14.47 ± 1.90	7.57 ± 1.93	10.30 ± 1.12
		500	16.34 ± 3.20	17.17 ± 1.52	17.11 ± 1.75	18.39 ± 2.13	15.54 ± 1.53	11.50 ± 1.34	10.63 ± 1.41
	shoot	0	17.38 ± 2.70	16.85 ± 1.68	16.24 ± 1.45	13.59 ± 2.51	8.42 ± 1.24	4.89 ± 0.56	4.42 ± 1.06
		200	11.41 ± 1.62	12.61 ± 0.96	11.92 ± 0.78	11.66 ± 2.31	7.47 ± 1.08	4.21 ± 0.39	5.28 ± 1.35
		500	8.07 ± 3.01	8.61 ± 1.34	9.44 ± 1.18	8.47 ± 1.95	6.89 ± 0.89	5.38 ± 1.15	4.73 ± 0.72

465

466 الم للاستشارات									
ił		Difference in	average root/shoo	t elongation (mea	Table 5 n, mm) of radish u	ınder joint pollutic	on of sulfamonome	ethoxine and cadm	ium
		Cd (mg/kg)			Sulfamonom	ethoxine(mg/kg) (Mean \pm SD)		
			0	0.5	1	2	4	8	16
	root	0	40.57 ± 4.13	41.90 ± 2.37	40.10 ± 3.59	35.12 ± 3.12	22.92 ± 2.91	14.59 ± 1.56	10.63 ± 0.98
		200	33.20 ± 3.26	32.69 ± 1.98	40.76 ± 4.01	29.35 ± 2.17	22.15 ± 1.46	14.10 ± 1.24	10.81 ± 1.01
		500	20.57 ± 1.75	27.53 ± 2.04	22.31 ± 2.56	23.91 ± 1.27	18.65 ± 2.42	17.23 ± 1.95	14.39 ± 1.25
	shoot	0	21.51 ± 2.31	20.71 ± 1.56	20.69 ± 0.91	21.59 ± 1.78	13.71 ± 1.24	9.70 ± 1.19	5.79 ± 0.99
		200	20.57 ± 2.42	17.00 ± 2.01	20.19 ± 1.21	15.91 ± 1.42	15.29 ± 1.51	8.45 ± 0.91	6.34 ± 1.95
		500	15.89 ± 1.95	16.43 ± 1.78	12.78 ± 1.56	12.12 ± 2.19	11.88 ± 0.95	8.48 ± 1.34	6.22 ± 1.05

www.m

Crop Species	Cd^{a}	Regression Equations ^b	R^2	Р	$\mathrm{IC}_{50}{}^{\mathrm{c}}$
Wheat	0	$RI = 0.8709C_{SMM} + 20.657$	0.8188	< 0.05	33.69
	200	$RI = 0.8861C_{SMM} + 27.663$	0.8854	< 0.01	25.21
	500	$RI = 0.5597C_{SMM} + 45.511$	0.9404	< 0.01	8.02
Chinese cabbage	0	$RI = 8.5443C_{SMM} + 12.331$	0.9263	< 0.01	4.41
	200	$RI = 6.1668C_{SMM} + 26.301$	0.8439	< 0.01	3.84
	500	$RI = 2.8312C_{SMM} + 42.666$	0.8049	< 0.01	2.59
Radish	0	$RI = 5.0153C_{SMM} + 5.7707$	0.8122	< 0.05	8.82
	200	$RI = 4.2274C_{SMM} + 15.847$	0.7903	< 0.01	8.08
	500	$RI = 2.0763C_{SMM} + 36.297$	0.7229	< 0.05	6.60

 Table 6

 Relationships between inhibitory rate of root elongation (RI) and the concentration of SMM at the same concentration of added Cd

^aThe concentration of added Cd, mg/kg.

^bC_{SMM} was the tested concentration of SMM, mg/kg.

^cIC₅₀ was the concentration of SMM corresponding to 50% inhibitory rate of RI, mg/kg.

and Cd to plants was less than each of them. This fact demonstrated that, in soil conditions, as an organic pollutant, SMM could change the surface characteristics of the soil by adsorption to the constituent soil, such as organic matter and clay, which made the soil adsorb more Cd and then reduce its bioavailability (Liu *et al.*, 2009). In addition, this may be related to the interactive effects of the two pollutants. SMM and Cd would form precipitation of Cd when the concentration of Cd was high in the solution, which would reduce the bioavailability of Cd and SMM. Nevertheless, the joint effects of chemicals and heavy metals could be determined by the interactive mode and the ratio of the two substances (Sun *et al.*, 2009).

Table 7

Relationships between inhibitory rate of shoot elongation (SI) and the concentration of SMM at the same concentration of added Cd

Crop Species	Cd^a	Regression Equations ^b	R^2	Р	IC ₅₀ ^c
Wheat	0	$SI = 1.1012C_{SMM} + 5.9302$	0.9211	< 0.01	40.02
Chinese cabbage	200	$SI = 0.8998C_{SMM} + 22.708$	0.6836	< 0.05	30.33
-	500	$SI = 0.2855C_{SMM} + 58.022$	0.7235	< 0.05	a
Radish	0	$SI = 10.491C_{SMM} + 0.6209$	0.9461	< 0.01	4.71
	200	$SI = 6.1868C_{SMM} + 27.936$	0.8717	< 0.01	3.57
	500	$SI = 3.136C_{SMM} + 47.963$	0.8769	< 0.01	0.65
	0	$SI = 4.8835C_{SMM} + 2.9261$	0.8732	< 0.01	9.64
	200	$SI = 4.1065C_{SMM} + 12.598$	0.8616	< 0.01	9.11
	500	$SI = 2.7276C_{SMM} + 32.066$	0.8452	< 0.01	6.58

^a The concentration of added Cd, mg/kg[.]

^b C_{SMM} was the tested concentration of SMM, mg/kg.

^c IC₅₀ was the concentration of SMM corresponding to 50% inhibitory rate of RI, mg/kg.



These results will help to further understand the phytotoxicity of an important antibiotic (SMM) and a common heavy metal (Cd), and their joint toxic effects. Future studies should be directed to phytotoxicity mechanisms; for example, possible uptake and translocation of antibiotics by plants, and the physical and chemical properties in the rhizosphere and on the root surface.

Conclusion

There were significant (P < 0.05) positive relationships between root and shoot elongation of the three crops (wheat, Chinese cabbage, and radish) and concentrations of the two pollutants (SMM and Cd), but seed germination of the three crops is not sensitive to the toxicity of the two pollutants. As far as root elongation was concerned, Chinese cabbage was the most sensitive to the toxicity of SMM and wheat was the most sensitive to the toxicological effects of Cd. Phytotoxicity of SMM on the three crops was much stronger than that of Cd. In the joint effects of SMM and Cd, the interactive effects of the two pollutants on root and shoot elongation of the three crops were very complicated. At the low concentration of SMM, SMM and Cd had significantly (P < 0.05) synergic effects on root and shoot elongation of the three crops. However, the synergic effects were not significant (P > 0.05), and antagonistic effects were found when the concentration of SMM was high.

Funding

The authors are grateful to the Henan Key Laboratory of Environmental Pollution Control and Key Laboratory for Yellow River and HuaiHe River Water Environmental and Pollution Control Ministry of Education. This work has been supported by the National Natural Science Foundation of China (No. 21107023 and No. 31170478), the China Postdoctoral Science Foundation funded project (No. 20110491001), and The Excellent Young Scientist Foundation of Shandong Province (No. 2010BSE27189).

References

- Accinelli, C., Koskinen, W. C., Becker, J. M. and Sadowsky, M. J. 2007. Environmental fate of two sulfonamide antimicrobial agents in soil. J. Agric. Food. Chem. 55, 2677–2682.
- Aina, R., Labra, M., Fumagalli, P., Vannini, C., Marsoni, M., Cucchi, U., Bracale, M., Sgorbati, S. and Citterio, S. 2007. Thiol-peptide level and proteomic changes in response to cadmium toxicity in *Oryza sativa L.* roots. *Environ. Exp. Bot.* **59**, 381–392.
- Baran, W., Sochacka, J., and Wardas, W. 2006. Toxicity and biodegradability of sulfonamides and products of their photocatalytic degradation in aqueous solutions. *Chemosphere*. 65, 1295–1299.
- Boxall, A. B., Blackwell, P., Cavallo, R., Kay, P., and Tolls, J. 2002. The sorption and transport of a sulphonamide antibiotic in soil systems. *Toxicol. Lett.* **131**, 19–28.
- Boxall, A. B., Fogg, L. A., Kay, P., Blackwell, P. A., Pemberton, E. J., and Croxford, A. 2003. Prioritisation of veterinary medicines in the UK environment. *Toxicol. Lett.* 142(3), 207–218.
- Cheng, Y. and Zhou, Q. X. 2002. Ecological toxicity of reactive X-3B red dye and cadmium acting on wheat (Triticum aestivum). *J. Environ. Sci.* **14**, 136–140.
- Christian, T., Schneider, R. J., Farber, H. A., Skutlarek, D., Meyer, M. T., and Goldbach, H. E. 2003. Determination of antibiotic residues in manure, soil, and surface waters. *Acta. Hydrochim. Hydrobiol.* 31, 36–44.



- Dagnac, T., Jeannot, R., and Mouvet, C. 2002. Determination of oxanilic and sulfonic acid metabolites of acetochlor in soils by liquid chromatography-electrospray ionization mass spectrometry. J. Chromatogr. A. 957, 69–77.
- Davis, J. G., Truman, C. C., Kim, S. C., Ascough, J. C., and Carlson, K. 2006. Antibiotic transport via runoff and soil loss. J. Environ. Qual. 35, 2250–2260.
- Dolliver, H., Kumar, K., and Gupta, S. 2007. Sulfamethazine uptake by plants from manure amended soil. J. Environ. Qual. 36, 1224–1230.
- Haller, M. Y., Müller, S. R., McArdell, C. S., Alder, A. C., and Suter, M. J. F. 2002. Quantification of veterinary antibiotics (sulfonamides and trimethoprim) in animal manure by liquid chromatography–mass spectrometry. J. Chromatogr. A. 952, 111–120.
- Hamscher, G., Pawelzick, H. T., Hoper, H., and Nau, H. 2005. Different behavior of tetracyclines and sulfonamides in sandy soils after repeated fertilization with liquid manure. *Environ. Toxicol. Chem.* 24, 861–868.
- He, Z. L., Zhou, Q. X., and Xie, Z. M. 1998. Soil-Chemical Equilibriums of Polluted and Beneficial Elements, pp. 115–120, Science Press, Beijing (in Chinese).
- Holger, H., Andreas, F., Marc, L., Kornelia, S., Michael, M., and Michael, S. 2008. Fate of sulfadiazine administered to pigs and its quantitative effect on the dynamics of bacterial resistance genes in manure and manured soil. *Soil. Biol. Biochem.* 40, 1892–1900.
- Ingerslev, F. and Halling-Srensen, B. 2000. Biodegradability properties of environmental chemistry: Biodegradability properties of sulfonamides in activated sludge. *Environ. Toxicol. Chem.* 19, 2467–2473.
- Jørgensen, S. E. and Halling-Sørensen, B. 2000. Drugs in the environment. Chemosphere. 40, 691-699.
- Klaus, K. 2009a. Antibiotics in the aquatic environment: A review: Part I. Chemosphere. 75, 417–434.
- Klaus, K. 2009b. Antibiotics in the aquatic environment: A review: Part II. Chemosphere. 75, 435-441.
- Kong, W. D. and Zhu, Y. G. 2007. A review on ecotoxicology of veterinary pharmaceuticals to plants and soil microbes. *Asian. J. Ecotoxicol.* 2, 1–9 (in Chinese).
- Lin, D. H. and Xing, B. S. 2007. Phytotoxicity of nanoparticles: Inhibition of seed germination and root growth. *Environ. Pollut.* 150, 243–250.
- Lin, R. Z., Wang, X. R., Luo, Y., Guo H. Y., and Yin D. Q. 2007. Effects of soil cadmium on growth, oxidative stress and antioxidant system in wheat seedlings (*Triticum aestivum L.*). *Chemosphere.* 69, 89–98.
- Liu, T. F., Wang, T., Sun, C., and Wang, Y. M. 2009. Single and joint toxicity of cypermethrin and copper on Chinese cabbage (Pakchoi) seeds. *J. Hazard. Mater.* **163**, 344–348.
- Malgorzata, W. and Jolanta, O. 1998. The effect of lead on seed imbibition and germination in different plant species. *Plant. Sci.* 137, 155–171.
- Nicole, K. 2008. Veterinary antibiotics in the aquatic and terrestrial environment. Ecol. Indic. 8, 1–13.
- Robert, S. and Joanna, D. 2003. Cadmium-induced changes in growth and cell cycle gene expression in suspension-culture cells of soybean. *Plant. Physiol. Biochem.* **41**, 767–772.
- Singh, O. V., Labana, S., Pandey, G., and Budhiraja, R. 2003. Phytoremediation: An overview of metallic ion decontamination from soil. *Appl. Microbol. Biotechnol.* 61, 405–412.
- Song, Y. F., Zhou, Q. X., Xu, H. X., Ren, L. P., and Gong, P. 2002. Ecological toxicity of heavy metals in soils acting on seed germination and root elongation of wheat. *Chin. J. Appl. Ecol.* 13, 459–462 (in Chinese).
- Sun, Y. B., Zhou, Q. X., Liu, W. T., An, J., Xu, Z. Q., and Wang, L. 2009. Joint effects of arsenic and cadmium on plant growth and metal bioaccumulation: A potential Cd-hyperaccumulator and As-excluder Bidens pilosa L. J. Hazard. Mater. 165(1–3), 1023–1028.
- Thiele-Bruhn, S. 2003. Pharmaceutical antibiotic compounds in soils: A review. J. Plant. Nutr. Soil. Sci. 166, 145–167.
- Thiele-Bruhn, B. S., Seibicke, T., Schulten, H. R., and Leinweber, P. 2004. Sorption of sulfonamide pharmaceutical antibiotics on whole soils and particle-size fractions. *J. Environ. Qual.* 33, 1331–1342.
- Wang, M. E. and Zhou, Q. X. 2006. Joint stress of chlorimuron-ethyl and cadmium on wheat Triticum aestivum at biochemical levels. *Environ. Pollut.* 144, 572–580.



- Wang, X. D., Sun, C., Wang, Y., and Wang, L. S. 2002. Quantitative structure-activity relationships for the inhibition toxicity to root elongation of Cucumis sativus of selected phenols and interspecies correlation with Tetrahymena pyriformis. *Chemosphere*. 46(2), 153–161.
- Wei, R. C., Ge, F., Huang, S. Y., Chen, M., and Wang, R. 2011. Occurrence of veterinary antibiotics in animal wastewater and surface water around farms in Jiangsu Province, China. *Chemo-sphere* 82(10), 1408–1414.
- Wojciech, B., Jolanta, S., and Wladyslaw, W. 2006. Toxicity and biodegradability of sulfonamides and products of their photocatalytic degradation in aqueous solutions. *Chemosphere*. 65, 1295–1299.
- Zhou, Q. X. 2003. Interaction between heavy metals and nitrogen fertilizers applied in soil– vegetable systems. *Bull. Environ. Contam. Toxicol.* **71**(2), 338–344.
- Zhou, Q. X., Kong, F. X., and Zhu, L. 2004a. *Ecotoxicology*, pp. 234–238, Science Press, Beijing (in Chinese).
- Zhou, Q. X., Cheng, Y., Zhang, Q. R., and Liang, J. D. 2004b. Quantitative analyses of relationships between ecotoxicological effects and combined pollution. *Sci. in. China.* **47**(4), 332–339.



Copyright of Soil & Sediment Contamination is the property of Taylor & Francis Ltd and its content may not be copied or emailed to multiple sites or posted to a listserv without the copyright holder's express written permission. However, users may print, download, or email articles for individual use.

